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=> s 11 and transport
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L5 2 L1 AND TRANSPORT

=> d ab 1-2

L5 ANSWER 1 OF 2 MEDLINE on STN

AB The Lyz endolysin of bacteriophage P1 was found to cause lysis of the host without a holin. Induction of a plasmid-cloned lyz resulted in lysis, and the lytic event could be triggered prematurely by treatments that dissipate the proton-motive force. Instead of requiring a holin, export was mediated by an N-terminal transmembrane domain (TMD) and required host sec function. Exported Lyz of identical SDS/PAGE mobility was found in both the membrane and periplasmic compartments, indicating that periplasmic Lyz was not generated by the proteolytic cleavage of the membrane-associated form. In gene fusion experiments, the Lyz TMD directed PhoA to both the membrane and periplasmic compartments, whereas the TMD of the integral membrane protein FtsI restricts Lyz to the membrane. Thus, the N-terminal domain of Lyz is both necessary and sufficient not only for export of this endolysin to the membrane but also for its release into the periplasm. The unusual N-terminal domain, rich in residues that are weakly hydrophobic, thus functions as a signal-arrest-release sequence, which first acts as a normal signal-arrest domain to direct the endolysin to the periplasm in membrane-tethered form and then allows it to be released as a soluble active enzyme in the periplasm. Examination of the protein sequences of related bacteriophage endolysins suggests that the presence of an N-terminal signal-arrest-release sequence is not unique to Lyz. These observations are discussed in relation to the role of holins in the control of host lysis by bacteriophage encoding a secretory endolysin.

L5 ANSWER 2 OF 2 MEDLINE on STN

AB Listeria monocytogenes bacteriophages A118, A500 and A511 are members of three distinct phage groups with characteristic host ranges. Their endolysin (ply) genes were cloned and expressed in Escherichia coli as demonstrated by the conferred lytic phenotype when colonies of recombinant cells were overlaid with a lawn of Listeria cells. The nucleotide sequences of the cloned DNA fragments were determined and the individual enzymes (PLY118, 30.8 kDa; PLY500, 33.4 kDa; PLY511, 36.5 kDa) were shown to have varying degrees of homology within their N-terminal or C-terminal domains. Transcriptional analysis revealed them to be 'late' genes with transcription beginning 15-20 min post-infection. The enzymes were overexpressed and partially purified and their individual specificities examined. When applied exogenously, the lysins induced rapid lysis of Listeria strains from all species but generally did not affect other bacteria. Using hydrolysis of purified listerial cell walls, PLY511 was characterized as an N-acetylmuramoyl-L-alanine amidase (EC 3.5.1.28) and shows homology in its N-terminal domain to other enzymes of this type. In contrast, PLY118 and PLY500 were shown to represent a new class of cell wall lytic enzymes which cleave between the L-alanine and D-glutamate residues of listerial peptidoglycan; these were designated as L-alanoyl-D-glutamate peptidases. These two enzymes share homology in the N-terminal domain which we propose determines hydrolytic specificity. Highly conserved holin (hol) gene sequences are present upstream of ply118 and ply500. They encode proteins of structural similarity to the product of phage lambda gene S, and are predicted to be membrane proteins which form pores to allow access of the lysins to their peptidoglycan substrates. This arrangement of conserved holin genes with downstream lysin genes among the siphoviral lysis cassettes explains why the cytoplasmic endolysins alone are not lethal, since they require a specific transport function across the cell membrane.

=> s 11 and antibacterial

24774 ANTIBACTERIAL
L6 1 L1 AND ANTIBACTERIAL

=> d ab

L6 ANSWER 1 OF 1 MEDLINE on STN

AB To characterize the enzymatic activity and antibacterial activity of endolysin encoded by a *Bacillus amyloliquefaciens* phage, the open reading frame encoding endolysin was amplified by PCR and cloned into the expression plasmid pET21d(+). The resultant plasmid was used to transform *Escherichia coli* JM109(DE3). Production of endolysin in the cytosol facilitated cell lysis without coproduction of holin, which is considered to degrade or alter the cytoplasmic membrane. The phage endolysin was overexpressed and purified. Although the specific activity of the purified phage endolysin towards lyophilized *Micrococcus luteus* cells was 1/11 of the activity of chicken egg white lysozymes, the endolysin showed stronger antibacterial activity towards *E. coli* W3110, *E. coli* JM109(DE3) and *Pseudomonas aeruginosa* PAO1 than chicken egg white lysozymes. The antibacterial activity of the endolysin towards these three bacterial strains was marked when EDTA was added to the endolysin solution.

=> s 11 and cell wall

2103676 CELL
156557 WALL
25927 CELL WALL
(CELL(W)WALL)

L7 19 L1 AND CELL WALL

=> d ab 1-19

L7 ANSWER 1 OF 19 MEDLINE on STN

AB The fate of phage-infected bacteria is determined by the holin, a small membrane protein that triggers to disrupt the membrane at a programmed time, allowing a lysozyme to attack the cell wall. S(21)68, the holin of phage 21, has two transmembrane domains (TMDs) with a predicted N-in, C-in topology. Surprisingly, TMD1 of S(21)68 was found to be dispensable for function, to behave as a SAR ("signal-anchor-release") domain in exiting the membrane to the periplasm, and to engage in homotypic interactions in the soluble phase. The departure of TMD1 from the bilayer coincides with the lethal triggering of the holin and is accelerated by membrane depolarization. Basic residues added at the N terminus of S(21)68 prevent the escape of TMD1 to the periplasm and block hole formation by TMD2. Lysis thus depends on dynamic topology, in that removal of the inhibitory TMD1 from the bilayer frees TMD2 for programmed formation of lethal membrane lesions.

L7 ANSWER 2 OF 19 MEDLINE on STN

AB Double-stranded DNA phages of both Gram-positive and Gram-negative bacteria typically use a holin-endolysin system to achieve lysis of their host. In this study, the lysis genes of *Staphylococcus aureus* phage P68 were characterized. P68 gene lys16 was shown to encode a cell-wall-degrading enzyme, which causes cell lysis when externally added to clinical isolates of *S. aureus*. Another gene, hol15, was identified embedded in the -1 reading frame at the 3' end of lys16. The deduced Hol15 protein has three putative transmembrane domains, and thus resembles class I holins. An additional candidate holin gene, hol12, was found downstream of the endolysin gene lys16 based on two predicted transmembrane domains of the encoded protein, which is a typical trait of class II holins. The synthesis of either Hol12 or Hol15 resulted in growth retardation of *Escherichia coli*, and

both hol15 and hol12 were able to complement a phage lambda Sam mutation. The hol15 gene has a dual start motif beginning with the codons Met1-Lys2-Met3.... Evidence is presented that the hol15 gene encodes a lysis inhibitor (anti-holin) and a lysis effector (actual holin). As depolarization of the membrane converted the anti-holin to a functional holin, these studies suggested that hol15 functions as a typical dual start motif class I holin. The unusual arrangement of the P68 lysis genes is discussed.

L7 ANSWER 3 OF 19 MEDLINE on STN

AB An open reading frame encoding an 88 amino acid protein was present downstream of the previously characterized endolysin of Streptomyces aureofaciens phage microl/6. Structural analysis of its sequence revealed features characteristic for holin. This open reading frame encoding the putative holin was amplified by polymerase chain reaction and cloned into the expression vector pET-21d(+). Synthesis of the holin-like protein resulted in bacterial cell death but not lysis. The holmicrol/6 gene was able to complement the defective lambda S allele in the nonsuppressing Escherichia coli HB101 strain to produce phage progeny. This fact suggests that the proteins encoded by both phage genes have analogous function, i.e. the streptomycete holin induces nonspecific lesions in the cytoplasmic membrane, through which the lambda endolysin gains an access to its substrate, the cell wall. The concomitant expression of both S. aureofaciens holmicro 1/6 and lambda endolysin in E. coli resulted in abrupt cell lysis. This result provided further evidence that the product of holmicro 1/6 gene is a holin.

L7 ANSWER 4 OF 19 MEDLINE on STN

AB An open reading frame homologous to the genes encoding several cell-wall hydrolyzing enzymes was identified on the genome of actinophage mu 1/6. This open reading frame encoding the putative endolysin was amplified by polymerase chain reaction and cloned into the expression vector PET-21a. This gene consisted of 1182 bp encoding a 393 amino acid polypeptide with a molar mass of 42.1 kDa. The gene product was overexpressed in Escherichia coli, and then the lytic enzyme was purified by a two-step chromatographic procedure. When applied exogenously, the endolysin of phage mu 1/6 was active against all tested Streptomyces strains but did not affect other bacteria. The amino acid sequence showed a high homology with a putative amidase of the Streptomyces phage phi C31. Downstream of the endolysin gene, an open reading frame encoding an 88 amino acid protein was identified. Structural analysis of its sequence revealed features characteristics for holin.

L7 ANSWER 5 OF 19 MEDLINE on STN

AB AIMS: To evaluate the ability of a filamentous phage encoding lethal proteins to kill bacteria without host-cell lysis. METHODS AND RESULTS: Bacterial survival was determined after infection of a growing Escherichia coli culture with phage M13 encoding either the restriction endonuclease BglII gene or modified phage lambda S holin genes. The genetically engineered phage exerted a high killing efficiency while leaving the cells structurally intact. When compared with a lytic phage, the release of endotoxin was minimized after infection with the genetically modified phages. CONCLUSIONS: Genetically engineered phage can be used for efficient killing, concomitantly minimizing endotoxin release. SIGNIFICANCE AND IMPACT OF THE STUDY: This feasibility study provides a possible strategy for the use of genetically engineered phage as bactericidal agents by optimizing the advantages and minimizing potential risks such as release of pyrogenic cell wall components.

L7 ANSWER 6 OF 19 MEDLINE on STN

AB Bacteriophages must destroy the bacterial cell wall to lyse their host and release their progeny into the environment. There are at least two distinct mechanisms by which phages destroy the cell wall. Bacteriophages with large genomes use a holin-endolysin system, while bacteriophages with small genomes encode a single lysis protein. Three unrelated single protein lysis systems are known and these proteins will be the focus of the review. Recent results indicate that at least two of these proteins inhibit cell wall synthesis and are thus the phage analogs of antibiotics like penicillin.

L7 ANSWER 7 OF 19 MEDLINE on STN

AB Clostridium perfringens commonly occurs in food and feed, can produce an enterotoxin frequently implicated in food-borne disease, and has a substantial negative impact on the poultry industry. As a step towards new approaches for control of this organism, we investigated the cell wall lysis system of *C. perfringens* bacteriophage phi3626, whose dual lysis gene cassette consists of a holin gene and an endolysin gene. Hol3626 has two membrane-spanning domains (MSDs) and is a group II holin. A positively charged beta turn between the two MSDs suggests that both the amino terminus and the carboxy terminus of Hol3626 might be located outside the cell membrane, a very unusual holin topology. Holin function was experimentally demonstrated by using the ability of the holin to complement a deletion of the heterologous phage lambda S holin in lambda delta Stf. The endolysin gene ply3626 was cloned in *Escherichia coli*. However, protein synthesis occurred only when bacteria were supplemented with rare tRNA(Arg) and tRNA(Ile) genes. Formation of inclusion bodies could be avoided by drastically lowering the expression level. Amino-terminal modification by a six-histidine tag did not affect enzyme activity and enabled purification by metal chelate affinity chromatography. Ply3626 has an N-terminal amidase domain and a unique C-terminal portion, which might be responsible for the specific lytic range of the enzyme. All 48 tested strains of *C. perfringens* were sensitive to the murein hydrolase, whereas other clostridia and bacteria belonging to other genera were generally not affected. This highly specific activity towards *C. perfringens* might be useful for novel biocontrol measures in food, feed, and complex microbial communities.

L7 ANSWER 8 OF 19 MEDLINE on STN

AB Holins are integral membrane proteins that control the access of phage-encoded muralytic enzymes, or endolysins, to the cell wall by the sudden formation of an uncharacterized homo-oligomeric lesion, or hole, in the membrane, at a precisely defined time. The timing of lambda-infected cell lysis depends solely on the 107 codon S gene, which encodes two proteins, S105 and S107, which are the holin and holin inhibitor, respectively. Here we report the results of biochemical and genetic studies on the interaction between the holin and the holin inhibitor. A unique cysteine at position 51, in the middle of the second transmembrane domain, is shown to cause the formation of disulfide-linked dimers during detergent membrane extraction. Forced oxidation of membranes containing S molecules also results in the formation of covalently linked dimers. This technique is used to demonstrate efficient dimeric interactions between S105 and S107. These results, coupled with the previous finding that the timing of lysis depends on the excess of the amount of S105 over S107, suggest a model in which the inhibitor functions by titrating out the effector in a stoichiometric fashion. This provides a basis for understanding two evolutionary advantages provided by the inhibitor system, in which the production of the inhibitor not only causes a delay in the timing of lysis, allowing the assembly of more virions, but also increases effective hole formation after triggering.

L7 ANSWER 9 OF 19 MEDLINE on STN

AB The genes encoding the host cell wall-lytic proteins were searched in the genome DNA of phage PL-1 active against Lactobacillus casei ATCC 27092 by comparing the amino acid sequences with those of others using a computer software of the DDBJ data base. The gene regions found were cloned into E. coli by inserting PCR-amplified DNA fragments into the EcoRI site of pUC 19, and the nucleotide sequences were determined. One of the ORFs (hol) consisted of 270 bp encoding 90 amino acids. The hol product (holin) possessed a putative secretion signal, two putative transmembrane helices, and a highly charged C-terminus. Another ORF (lys) consisted of 1050 bp encoding an N-acetylmuramoyl-L-alanine amidase of 350 amino acids. The gene lys was expressed in E. coli using pCALn expression vector, and the purified gene product hydrolysed the amide linkage in the peptidoglycans of L. casei. The amino acid sequence of PL-1 amidase showed a high homology to those of Lactococcus lactis phage rlt and Listeria monocytogenes phage A511. It was suggested that the N-terminal region was involved in enzyme activity and the C-terminal region in binding the enzyme to the cell wall substrate, respectively.

L7 ANSWER 10 OF 19 MEDLINE on STN

AB We have cloned, sequenced, and characterized the genes encoding the lytic system of the unique Staphylococcus aureus phage 187. The endolysin gene ply187 encodes a large cell wall-lytic enzyme (71.6 kDa). The catalytic site, responsible for the hydrolysis of staphylococcal peptidoglycan, was mapped to the N-terminal domain of the protein by the expression of defined ply187 domains. This enzymatically active N terminus showed convincing amino acid sequence homology to an N-acetylmuramoyl-L-alanine amidase, whereas the C-terminal part, whose function is unknown, revealed striking relatedness to major staphylococcal autolysins. An additional reading frame was identified entirely embedded out of frame (+1) within the 5' region of ply187 and was shown to encode a small, hydrophobic protein of holin-like function. The hol187 gene features a dual-start motif, possibly enabling the synthesis of two products of different lengths (57 and 55 amino acids, respectively). Overproduction of Hol187 in Escherichia coli resulted in growth retardation, leakiness of the cytoplasmic membrane, and loss of de novo ATP synthesis. Compared to other holins identified to date, Hol187 completely lacks the highly charged C terminus. The secondary structure of the polypeptide is predicted to consist of two small, antiparallel, hydrophobic, transmembrane helices. These are supposed to be essential for integration into the membrane, since site-specific introduction of negatively charged amino acids into the first transmembrane domain (V7D G8D) completely abolished the function of the Hol187 polypeptide. With antibodies raised against a synthetic 18-mer peptide representing a central part of the protein, it was possible to detect Hol187 in the cytoplasmic membrane of phage-infected S. aureus cells. An important indication that the protein actually functions as a holin in vivo was that the gene (but not the V7D G8D mutation) was able to complement a phage lambda Sam mutation in a nonsuppressing E. coli HB101 background. Plaque formation by lambdagt11::hol187 indicated that both phage genes have analogous functions. The data presented here indicate that a putative holin is encoded on a different reading frame within the enzymatically active domain of ply187 and that the holin is synthesized during the late stage of phage infection and found in the cytoplasmic membrane, where it causes membrane lesions which are thought to enable access of Ply187 to the peptidoglycan of phage-infected Staphylococcus cells.

L7 ANSWER 11 OF 19 MEDLINE on STN

AB Upon infecting populations of susceptible host cells, T-even bacteriophages maximize their yield by switching from lysis at about 25 to 35 min at 37 degrees C after infection by a single phage

particle to long-delayed lysis (lysis inhibition) under conditions of sequential infection occurring when free phages outnumber host cells. The timing of lysis depends upon gene t and upon one or more rapid-lysis (r) genes whose inactivation prevents lysis inhibition. t encodes a holin that mediates the movement of the T4 endolysin through the inner cell membrane to its target, the cell wall. The rI protein has been proposed to sense superinfection. Of the five reasonably well characterized r genes, only two, rI and rV, are clearly obligatory for lysis inhibition. We show here that rV mutations are alleles of t that probably render the t protein unable to respond to the lysis inhibition signal. The tr alleles cluster in the 5' third of t and produce a strong r phenotype, whereas conditional-lethal t alleles produce the classical t phenotype (inability to lyse) and other t alleles produce additional, still poorly understood phenotypes. tr mutations are dominant to t+, a result that suggests specific ways to probe T4 holin function.

L7 ANSWER 12 OF 19 MEDLINE on STN

AB Holins are small membrane proteins that, at a genetically programmed time in a bacteriophage infective cycle, allow bacteriolytic enzymes, or endolysins, to escape to the periplasm and to attack the cell wall. Most holins fall into two sequence classes, I and II, based on the number of potential transmembrane domains (three for class I and two for class II). The prototype class I holin gene, S lambda, has a dual start motif and encodes not only the effector holin, Slambda105, but also an inhibitor, Slambda107, with a Met-Lys ...extension at the terminus. The prototype class II holin gene of phage 21, S 21, begins with the motif Met-Lys-Ser-Met ..., and a potential RNA secondary structure overlaps the Shine-Dalgarno sequence. Here, we demonstrate that (i) two protein products are elaborated from S 21, S2171 and S2168; (ii) the shorter product is required for lysis; (iii) the longer product, S2171, inhibits S 21 function; and (iv) the Lys-2 residue is important for the inhibitor function. Moreover, the RNA stem-loop structure is involved in the downregulation of S2171 synthesis. However, our results suggest that, in S 21, different segments of the single consensus Shine-Dalgarno sequence serve the two translational starts. These results show that the dual start motifs of class II holin genes are functionally homologous to those of class I holin genes.

L7 ANSWER 13 OF 19 MEDLINE on STN

AB The lysis genes of the virulent *Staphylococcus aureus* bacteriophage Twort were cloned and their nucleotide sequences determined. The endolysin gene plyTW encodes a 53.3-kDa protein, whose catalytic site is located in the amino-terminal domain. An enzymatically active fragment (N-terminal 271 amino acids) was overexpressed in *Escherichia coli* and partially purified. The enzyme rapidly cleaves staphylococcal peptidoglycan, and was shown to act as N-acetylmuramoyl-L-alanine amidase (EC 3.5.1.28). Significant sequence homology to the specific cell wall targeting domain of lysostaphin was observed in a 101-amino acid C-terminal overlap. However, we found that the large C-terminal portion (63%, 295 aa) of PlyTW is not required for staphylolytic activity. Located upstream of and overlapping plyTW by 35 bp in a different reading frame (+1), we identified holtW, which starts with a single TTG triplet. The gene specifies a 185-amino acid (20.5 kDa) holin protein, which features two potential hydrophobic, antiparallel transmembrane domains, and a highly charged, acidic C-terminus. HoltW is the largest class II holin described to date. It can substitute for the defective allele in phase lambda S' amber mutants, both in trans from an expression plasmid, and from within gt11::holTW. The proposed function is the formation of unspecific membrane lesions to promote access of the endolysin to the bacterial peptidoglycan.

L7 ANSWER 14 OF 19 MEDLINE on STN

AB Lactobacillus plantarum phage phi gle has two consecutive cell lysis genes hol-lys (Oki et al., 1996b). In the present study, functional and structural properties of the hol protein (Hol) were characterized in Escherichia coli. Electron microscopic examinations showed that hol under plac in E. coli XL1-Blue injured the inner membrane to yield empty ghost cells with the bulk of the cell wall undisturbed. Northern blot analysis indicated that hol-lys genes under plac were co-transcribed, although the amount of hol transcript was larger than that of lys, ceasing via an apparently rho-independent terminator just downstream of hol. However, deletion and/or fusion experiments suggested that: (1) the N-terminal half of phi gle Hol composed of three putative transmembrane domains may be responsible for interaction with membrane; (2) the N-terminal end (five amino acids) seems nonessential; and (3) the C-terminal half containing charged amino acids appears to be involved in proper hol function. These results suggest that phi gle Hol is a member of the lambdoid holin family, but divergent in several properties from lambda holin.

L7 ANSWER 15 OF 19 MEDLINE on STN

AB We have characterized four families of pneumococcal phages with remarkable morphologic and physiological differences. Dp-1 and Cp-1 are lytic phages, whereas HB-3 and EJ-1 are temperate phages. Interestingly, Cp-1 and HB-3 have a terminal protein covalently linked to the 5' ends of their linear DNAs. In the case of Dp-1, we have found that the choline residues of the teichoic acid were essential components of the phage receptors. We have also developed a transfection system using mature DNAs from Dp-4 and Cp-1. In the later case, the transfecting activity of the DNA was destroyed by treatment with proteolytic enzymes, a feature also shared by the genomes of several small Bacillus phages. DNA replication was investigated in the case of Dp-4 and Cp-1 phages. The terminal protein linked to Cp-1 DNA plays a key role in the peculiar mechanism of DNA replication that has been coined as protein-priming. Recently, the linear 19,345-bp double-stranded DNA of Cp-1 has been completely sequenced, several of its gene products have been analyzed, and a complete transcriptional map has been elaborated. Most of the pneumococcal lysins exhibit an absolute dependence of the presence of choline in the cell wall substrate for activity, and phage lysis requires, as reported for other systems, the action of a second phage-encoded protein, the holin, which presumably forms some kind of lesion in the membrane. The two lytic gene cassettes, from EJ-1 and Cp-1 phages, have been cloned and expressed in heterologous and homologous systems. The finding that some lysogenic strains of Streptococcus pneumoniae harbor phage remnants has provided important clues on the interchanges between phage and bacteria and supports the view of the chimeric origin of phages.

L7 ANSWER 16 OF 19 MEDLINE on STN

AB The ply genes encoding the endolysin proteins from *Bacillus cereus* phages Bastille, TP21, and 12826 were identified, cloned, and sequenced. The endolysins could be overproduced in *Escherichia coli* (up to 20% of total cellular protein), and the recombinant proteins were purified by a two-step chromatographical procedure. All three enzymes induced rapid and specific lysis of viable cells of several *Bacillus* species, with highest activity on *B. cereus* and *B. thuringiensis*. Ply12 and Ply21 were experimentally shown to be N-acetylmuramoyl-L-alanine amidases (EC 3.5.1.28). No apparent holin genes were found adjacent to the ply genes. However, Ply21 may be endowed with a signal peptide which could play a role in timing of cell lysis by the cytoplasmic phage endolysin. The individual lytic enzymes (PlyBa, 41.1 kDa; Ply21, 29.5 kDa, Ply12, 27.7 kDa) show remarkable heterogeneity, i.e., their amino acid sequences reveal only little homology. The N-terminal part of Ply21 was found to be almost identical to the catalytic domains of a *Bacillus* sp. cell wall hydrolase (CwlSP) and an autolysin of *B. subtilis* (CwlA). The C terminus of PlyBa contains a 77-amino-acid

sequence repeat which is also homologous to the binding domain of CwlSP. Ply12 shows homology to the major autolysins from *B. subtilis* and *E. coli*. Comparison with database sequences indicated a modular organization of the phage lysis proteins where the enzymatic activity is located in the N-terminal region and the C-termini are responsible for specific recognition and binding of *Bacillus* peptidoglycan. We speculate that the close relationship of the phage enzymes and cell wall autolysins is based upon horizontal gene transfer among different *Bacillus* phages and their hosts.

L7 ANSWER 17 OF 19 MEDLINE on STN

AB The lysis genes of a *Lactobacillus* phage phi gle were cloned, sequenced, and expressed in *Escherichia coli*. Nucleotide sequencing of a 3813-bp phi gle DNA revealed five successive open reading frames (ORF), Rorf50, Rorf118, hol, and lys and Rorf175, in the same DNA strand. By comparative analysis of the DNA sequence, the putative hol product (holin) has an estimated molecular weight is 14.2 kDa, and contains two potential transmembrane helices and highly charged N- and C-termini, resembling predicted holins (which are thought to be a cytoplasmic membrane-disrupting protein) encoded by other phages such as mv1 from *Lactobacillus bulgaricus*, phi adh from *Lactobacillus gasseri*, as well as monocins from *Listeria*. On the other hand, the putative phi gle lys product (lysin) of 48.4 kDa shows significant similarity with presumed muramidase, known as a cell wall peptidoglycandegrading enzyme, encoded by the *Lactobacillus* phage mv1 and phi adh, the *Lactococcus lactis* phage phi LC3, and the *Streptococcus pneumoniae* phages Cp-1, Cp-7 and Cp-9. When expressed in *E. coli*, the phi gle lysis and/or holin decreased the cell turbidity significantly, suggesting that the phi gle hol-lys system is involved in cytolytic process.

L7 ANSWER 18 OF 19 MEDLINE on STN

AB The holin function Ejh of the pneumococcal bacteriophage EJ-1 has been characterized. It shows structural features similar to, and functionally complemented, the prototype member of the holin family. In *Escherichia coli* and *Pseudomonas putida* the Ejh product caused cellular death, and changes in cell morphology could be accounted for by lesions in the cytoplasmic membrane. Expression of ejh resulted in the inhibition of growth in a variety of phylogenetically distant bacterial genera, suggesting a broad spectrum of action. Concomitant expression of the ejh and ejl (encodes a lysis) genes led to lysis of *E. coli* and *P. putida* cells. Remarkably, the Ejl lysis was able to attack murein from bacteria lacking choline in their sacculi, which suggests that pneumococcal lysins have a broader substrate specificity than previously assumed. Furthermore, the Ejh holin was able to trigger activity of the major pneumococcal autolysin cloned and expressed in *E. coli*, and this raised new questions about the regulation of this model autolysin. A new function for holins in systems where the phage lysis is supposed to be associated with the membrane is proposed.

L7 ANSWER 19 OF 19 MEDLINE on STN

AB *Listeria monocytogenes* bacteriophages A118, A500 and A511 are members of three distinct phage groups with characteristic host ranges. Their endolysin (ply) genes were cloned and expressed in *Escherichia coli* as demonstrated by the conferred lytic phenotype when colonies of recombinant cells were overlaid with a lawn of *Listeria* cells. The nucleotide sequences of the cloned DNA fragments were determined and the individual enzymes (PLY118, 30.8 kDa; PLY500, 33.4 kDa; PLY511, 36.5 kDa) were shown to have varying degrees of homology within their N-terminal or C-terminal domains. Transcriptional analysis revealed them to be 'late' genes with transcription beginning 15-20 min post-infection. The enzymes were overexpressed and partially purified and their individual specificities examined. When applied exogenously, the lysins induced rapid lysis of *Listeria* strains from all species but generally did not

affect other bacteria. Using hydrolysis of purified listerial cell walls, PLY511 was characterized as an N-acetylmuramoyl-L-alanine amidase (EC 3.5.1.28) and shows homology in its N-terminal domain to other enzymes of this type. In contrast, PLY118 and PLY500 were shown to represent a new class of cell wall lytic enzymes which cleave between the L-alanine and D-glutamate residues of listerial peptidoglycan; these were designated as L-alanoyl-D-glutamate peptidases. These two enzymes share homology in the N-terminal domain which we propose determines hydrolytic specificity. Highly conserved holin (hol) gene sequences are present upstream of ply118 and ply500. They encode proteins of structural similarity to the product of phage lambda gene S, and are predicted to be membrane proteins which form pores to allow access of the lysins to their peptidoglycan substrates. This arrangement of conserved holin genes with downstream lysin genes among the siphoviral lysis cassettes explains why the cytoplasmic endolysins alone are not lethal, since they require a specific transport function across the cell membrane.

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L7 ANSWER 6 OF 19 MEDLINE on STN
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CS Department of Molecular Biology and Microbiology, Case Western Reserve
University, School of Medicine, Cleveland, OH 44106, USA.
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